

**Research Paper**

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ISSN 0189-6016©2007**PHYTOCHEMICAL AND ANTIMICROBIAL EFFECTS OF *CHROZOPHORA SENEGALENSIS******Usman, H¹., Musa, Y. M²., Ahmadu, A. A³, Tijjani, M. A¹.**¹Department of Chemistry, University of Maiduguri, Maiduguri, Nigeria.²Department of Basic Sciences, College of Agriculture, Lafia, Nasarawa State, Nigeria.³Department of Pharm. & Medicinal Chem., Ahmadu Bello University, Zaria, Nigeria.**E-mail:** husman321@yahoo.com**Abstract**

The *in vitro* antimicrobial activities of the whole plant extract (ethanolic-CEE) of *Chrozophora senegalensis* and its fractions (ethyl acetate-EAA, n-butanol-NBE, aqueous-AQE) were assayed using the agar plate diffusion and nutrient broth dilution methods. Test microorganisms were *Bacillus subtilis* (NCTC 8326 B76), *Escherichia coli* (ATCC 11775), *Pseudomonas aeruginosa* (ATCC 10145), *Staphylococcus aureus* (ATCC 021001), *Aspergillus flavus*, *Aspergillus niger*, *Candida albicans* and *Salmonella typhi* - laboratory isolates. CEE, EAA and NBE inhibited all the test bacterial organisms and a fungus-*Aspergillus flavus*. AQE inhibited only *Salmonella typhi* and *Bacillus subtilis*. None of the extracts had activity on other 3 fungal organisms tested. CEE and EAA showed minimum inhibition concentration (MIC) of 0.390 and 3.125 mg/ml against *S. typhi* and *E. coli*, while NBE and AQE had MIC of 3.125 and 1.563 mg/ml against *S. typhi* respectively. NBE had an MIC of 12.500 mg/ml against *E. coli*. The minimum bactericidal concentration (MBC) of CEE and EAA was found to be <0.098 against *S. typhi*. The MBC of AQE was 12.5 mg/ml against *E. coli* and *S. aureus*, and 6.25 mg/ml towards *P. aeruginosa*. CEE and EAA exhibited similar antibacterial activities, followed by AQE. The extracts revealed the presence of carbohydrates, tannins, saponins, sterols determined by utilizing standard methods of analysis. This study has justified the traditional use of the plant for treating diarrhea, boils and syphilis.

Key words: Antimicrobial activity, *Chrozophora senegalensis*, Extracts, Phytochemical Screening, Euphorbiaceae.**Introduction**

Plant is man's friend in survival, giving him food and fuel, shelter and medicine from the days beyond the dawn of civilization (Bose and Choudhary, 1991). Although, the use of herbs as remedy for treatment of disease ailments has declined in the West, it continues to exist throughout the developing World (Rahman and Choudhary, 1999). Many African plants are used in traditional medicine as antimicrobial agents but only few are documented (Bellomaria and Kacou, 1995; Lewis and Elvin-Lewis, 1997; Ahmad, et al., 1998). However, over 80% of the world's population use plant as their primary source of medication (Cordell, 2000) and in view of the fact that antibiotics are sometimes associated with adverse side effects to the host including hypersensitivity, immuno-suppressive and allergic reactions, it is of interest to develop alternative antimicrobial drugs such as medicinal plants for the treatment of infectious diseases (Clark, 1996).

The plant *Chrozophora senegalensis* A. Juss (family: Euphorbiaceae) is herb with a deep-red flowers and violet tingled capsules; found common in dried-up inundated flats or sandy river beds (Dalziel, 1955). In Northern Nigeria, the plant is an astringent for diarrhea taken boiled with cereal foods and also to treat boils. A decoction is used as a body wash by pregnant women. It is chiefly used as a remedy for syphilis; also as an ingredient in a

mixture called “*Rigakafi*” – some other ingredients are roots and leaves of *Crotalaria spp.* *Portulaca Oleracea*, *Feretia Canthiothes*, *etc.* is taken as a reputed preventive and cure for the treatment of syphilis (Dalziel, 1955). In some districts, it is used as black dye. The plants of the genus *Chrozophora* have extra-floral nectarines; as one of the Hausa names suggests (Dalziel, 1955). The plant has been reported to serve as a remedy for intestinal pain, for conjunctivitis as well as cicatrizant (Tignokpa *et al.*, 1986; Etkin, 1997). The plant is known locally as: *Damagi*, *Damangi* or *Bauren kiyashi*, also as *Walkin machiji* in “Hausa” language (Dalziel, 1955).

There was no report on the extensive phytochemical studies of this plant species. Meanwhile, in this investigation, the antimicrobial effects of the crude ethanolic extract of the whole plant and its partitioned fractions against organisms commonly implicated in diseases reported earlier including *S. typhi* – the major causative agent for typhoid fever were reported. To the best of our knowledge, this is the first time the whole plant extracts was screened for antimicrobial activities.

Experimental

Plant Material

The whole plants of *Chrozophora senegalensis* A. Juss (voucher No. 579) were collected in April 2004, from Kudingi village, Samaru-Zaria, Kaduna State, Nigeria. The plant specimen was identified by Mr. U.A. Gallah and authenticated at the Herbarium in Biological Science Department, Ahmadu Bello University, Zaria – Nigeria, where a herbarium specimen was deposited.

Extraction and preparation of plant extract

The plant material (whole) was air-dried at room temperature for five days and pulverized with mortar and pestle. Four hundred grams of the powered plant material was defatted with petroleum ether (60-80°C) using continuous soxhlet extraction method to exhaustion. The extract was then concentrated *in vacuo* and greenish-yellow viscous oil 15.0 g (3.75% w/w) coded “PEE” was obtained. The resulting marc was exhaustively extracted by same method using 95% ethanol in water and a greenish-brown gummy mass 66.0 g (16.5% w/w) coded “CEE” was obtained after concentration *in vacuo*. About 35 g of the crude ethanolic extract (CEE) was completely suspended in 250 ml distilled water and then filtered. The resulting filtrate was then partitioned with ethyl acetate until the resulting organic layer was visibly clear, the ethyl acetate soluble portion was then concentrated *in vacuo* to generate a yellowish-brown mass 2.25 g (6.43% w/w) coded “EAA”; then the resulting aqueous layer partitioned with n-butanol until the resulting organic layer was visibly clear; the n-butanol soluble portion was also concentrated *in vacuo* to generate a dark-brown mass 10.01 g (28.6% w/w) coded “NBE”. The residual aqueous layer was finally concentrated *in vacuo* to yield a reddish-brown mass 14.18 g (40.5% w/w) coded “AQE”. The extracts were stored aseptically in a desiccator at room temperature until demanded.

Phytochemical screening

The crude ethanolic extract and its partitioned extracts were screened phytochemically for the presence of its constituents utilizing standard methods of analyses (Sofowora, 1993; Harborne, 1993; Trease and Evans, 2002).

Bacterial and Fungal strains

The Gram-positive organisms used in this study are *Bacillus subtilis* (NCTC 8326 B76) and *Staphylococcus aureus* (ATCC 021001). Gram-negative organisms are *Escherichia coli* (ATCC 11775), *Pseudomonas aeruginosa* (ATCC 10145) and *Salmonella typhi*. The fungal strains, *Candida albicans*, *Aspergillus flavus* and *Aspergillus niger*, are clinical isolates. All the organisms were obtained from the Department of Pharmaceutics and Pharmaceutical Microbiology, Ahmadu Bello University, Zaria – Nigeria.

Antimicrobial tests

Antimicrobial activities of the 4 extracts (CEE, EAA, NBE and AQE) were determined using disc diffusion technique as described by Odama, *et al.*, (1986); Chung, *et al.*, (1990), using a stock concentration of 100 mg/ml, prepared by dissolving 1 g of each extract into 10 ml of sterilized distilled water. The microorganisms were maintained on agar slants. The inocula were prepared by inoculating the test organisms in nutrient broth and incubating them for 24 hours at 37 °C while for *A. flavus* and *A. niger*, sabouraud dextrose broth was used which

was incubated for 48 hours. After incubation, the broth cultures were diluted to 1:1000 for Gram-positive bacteria and 1:5000 for the Gram-negative bacteria. One milliliter of the diluted cultures was inoculated into 19 ml sterile molten nutrient agar (48 °C) and poured into sterile petri dishes. Similarly, 1 ml of the diluted fungal suspensions was poured onto sterile Sabouraud dextrose agar plates and the excess sucked up with sterile Pasteur pipette. These were gently swirled and allowed to solidify. Afterwards, wells were bored into the solidified and inoculated nutrient agar plates using number 4 cork borer. All the wells were filled with equal volumes of 0.1 ml (10 mg/hole) of each extract. Ampicillin (25 µg), Chloramphenicol (10 µg), Ciprofloxacin (5 µg), Gentamicin (50 µg), Nalidixic acid (30 µg) standard discs were placed on the agar plate. About an hour was allowed for the extract to diffuse into the agar. Plates were then incubated overnight at 25 °C and 37 °C for fungi and bacterial strains respectively. At the end of the incubation period, inhibition zones were recorded in millimeters as the diameter of growth free zones around the bored holes using a transparent metre rule. Each extract and standard antibiotics were independently tested in duplicate. Diameters of zones of inhibition ≥ 10 mm were considered active (Zwadyk, 1972).

Minimum inhibitory concentration (MIC)

MIC was determined using the broth dilution technique as described by Sidney *et al.*, (1978); Vollekova *et al.*, (2001). The minimal inhibitory concentration value was determined for the microorganisms that were sensitive to the extracts under study (CEE, EAA, NBE and AQE). The microorganisms were prepared as described earlier. Each extract was first diluted to the highest concentration (100 mg/ml) in sterile distilled water, and then two-fold serial dilution of each extract was made to a concentration ranging from 0.098 - 50 mg/ml using nutrient broth. The extracts were inoculated 0.2 ml suspension of the organisms. MIC is defined as the lowest concentration where no visible turbidity was observed in the test tubes.

Minimum bactericidal concentration (MBC)

MBC were determined by using the broth dilution technique previously described by Vollekova *et al.*, (2001) by assaying the test tubes resulting from MIC determinations. A loopful of the content of each test tube was independently inoculated by streaking on a solidified nutrient agar plate incubated at 37°C for 24 hours and then observed for bacterial growth. The lowest concentration of the subculture with no growth was considered the minimum bactericidal concentration.

Results and Discussion

As shown in Table 1, the phytochemical screening of the crude ethanolic-extract (CEE) and its fractions [EAA, NBE and AQE] revealed the presence of carbohydrates, saponins, tannins and steroids. Alkaloids and flavonoids were not detected in the extracts of *Chrozophora senegalensis*. These classes of compounds are known to show curative activity against several pathogens (Hassan *et al.*, 2004) and therefore could explain its use traditionally for the treatment of wide array of illnesses.

The antimicrobial screenings are recorded in Table 2 expressing the zones of inhibition of bacterial and fungal growth. The extracts showed considerable amount of inhibition against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*; with much activity on *Salmonella typhi*. All the extracts had appreciable activities against *Aspergillus flavus* but none on other fungal organisms, *Candida albicans* and *Aspergillus niger*. The standard antibiotics disc used in this study inhibited the growth of the test bacteria but inactive against fungal organisms. The zones of inhibition produced by most antibiotic discs against *E. coli* and *S. typhi* were found to be smaller than those produced by some extracts especially CEE and EAA.

From the results of the MIC and MBC presented in Tables 3 and 4 respectively, it can be surmised that the active constituents responsible for the widest activities in this plant was residing in the mid-polar (EAA) fraction of the ethanolic extracts compared with other fractions (NBE and AQE); this was basically because the CEE and EAA fraction notably exhibited minimal inhibitory concentration of 0.390 and 3.125 mg/ml against the Gram-negative organisms (*S. typhi* and *E. coli*). NBE had an MIC of 12.500 mg/ml against *E. coli*. The minimum bactericidal concentration of ethanolic and ethyl acetate extracts was found to be <0.098 against *S. typhi*. For n-butanol extract, the MBC was 12.5 mg/ml against *E. coli* and *S. aureus*, and 6.25 mg/ml towards *P. aeruginosa*. The ethanolic and ethyl acetate extract exhibited similar antibacterial activities, followed by the n-butanol extract. The AQE had a good activity on *S. typhi*, giving 1.563 mg/ml as MIC and MBC.

All the extracts exhibited appreciable activity against *S. aureus*, a pyogenic bacterium known to play a significant role in invasive skin diseases including superficial and deep follicular lesion (Srinivasan, *et al.*, 2001). The broad-

spectrum antibacterial activity exhibited by the CEE and EAA could not be unrelated with the high concentrations of tannins, saponins and sterols in these extracts. In line with these findings, Trease and Evans, (1978) reported that tannins had been widely used as an application to sprains, bruises and superficial wounds. The inability of AQE to inhibit the growth of some Gram-negative organisms could be probably due to little or no much left from the exhaustive partitioning using the less polar solvents.

In conclusion, the fact that the extracts produced inhibitory activities against almost all the test bacteria and a fungus and marked higher activities than most of the reference drugs on the microbes provides some scientific basis for some of the uses in traditional medicine against some of the claim mentioned earlier. It is also of interest to report for the first time that extract from this plant could serve as a remedy for typhoid fever. This finding lends credence to the traditional use of the plant as a remedy for boils, diarrhea, syphilis and intestinal pain. The plant could be a potent anti-typhoid preparation.

We therefore, suggest the isolation and possible characterization of the active constituent(s) from the extracts of this plant species as possible antibacterial agents.

Table 1: Phytochemical analysis of the ethanolic extract of *Chrozophora senegalensis* and its fractions

S/No	Constituents/ Test	Extracts			
		CEE	EAA	NBE	AQE
1.	Alkaloids				
	Dragendorff's	-	-	-	-
	Mayer's	-	-	-	-
	Wagner's	-	-	-	-
2.	Cardenolides aglycon				
	Legal Test	++	+	++	+
	Keller-killiani Test	+++	++	+++	-
3.	Carbohydrates				
	Molisch's Test	+++	+	+	+++
	Barfoed's Test	+	-	-	++
	Fehling (reducing sugar)	+++	+	+	+++
	Fehling (combine reducing sugar)	++	+	-	+++
4.	Flavonoids				
	Shinoda's Test	-	-	-	-
	NaOH Test	-	-	-	-
5.	Saponins				
	Frothing Test	+++	+	++	+++
6.	Steroidal nucleus				
	Salkowski Test	++	+	++	+
	Liebermann – Burchard's Test	-	-	-	-
7.	Tannins				
	FeCl ₃ Test	+++	+	++	+
	Lead acetate Test	++	++	++	+

Key: CEE - Crude ethanolic extract, EAA - ethyl acetate portion, NBE - n-butanol soluble portion, AQE - residual aqueous portion. +++ = Highly present, ++ = moderately present, + = faintly present, - = absent.

Table 2: Susceptibility study of the extracts of *Chrozophora senegalensis* against test microorganisms

Extracts / Standards	Conc. (mg/hole)	Inhibition Zone (mm)							
		B.s	E.c	P.e	S.a	S.t	A.f	A.n	C.a
CEE	10	18.5 ± 0.5	22.5 ± 0.5	12.5 ± 0.5	21.0 ± 0.0	33.0 ± 0.0	11.5 ± 0.5	-	-
EAA	10	15.0 ± 0.0	30.5 ± 0.5	14.5 ± 0.5	20.5 ± 0.5	33.5 ± 0.5	13.5 ± 0.5	-	-
NBE	10	15.5 ± 0.5	20.0 ± 0.0	10.5 ± 0.5	19.5 ± 0.5	30.0 ± 0.0	12.5 ± 0.5	-	-
AQE	10	12.0 ± 0.0	-	-	-	22.5 ± 0.5	12.5 ± 0.5	-	-
AMP	25µg/disc	14.0 ± 0.0	14.0 ± 0.0	13.0 ± 0.0	10.0 ± 0.0	-	-	-	-
CHL	10µg/disc	ND	ND	12.0 ± 0.0	12.0 ± 0.0	12.0 ± 0.0	-	-	-
CIP	5µg/disc	24.0 ± 0.0	22.0 ± 0.0	22.0 ± 0.0	23.0 ± 0.0	16.0 ± 0.0	-	-	-
GEN	50µg/disc	20.0 ± 0.0	14.0 ± 0.0	14.0 ± 0.0	22.0 ± 0.0	16.0 ± 0.0	-	-	-
NAL	30µg/disc	14.0 ± 0.0	20.0 ± 0.0	14.0 ± 0.0	14.0 ± 0.0	18.0 ± 0.0	-	-	-
*Methanol	100%	-	-	-	-	-	-	-	-

Key: B.s = *Bacillus subtilis*, E.c = *Escherichia coli*, P.e = *Pseudomonas euroginosa*, S.a = *Staphylococcus aureus*, S.t = *Salmonella typhi*, C.a = *Candida albicans*, A.f = *Aspergillus flavus*, A.n = *Aspergillus niger*, CEE = crude ethanolic extract, EAA = ethyl acetate portion, NBE = n-butanol portion, AQE = residual aqueous portion, AMP = ampicillin, CHL = chloramphenicol, CIP = ciprofloxacin, GEN = gentimicin, NAL = nalidixic acid, ND = no disc, - = no inhibition, * = working solvent.

Table 3: Minimum inhibitory concentrations (MIC) of the extracts *Chrozophora senegalensis*

Extracts	Organisms	Concentration (mg/ml)									
		0.098	0.195	0.390	0.780	1.563	3.125	6.250	12.500	25.000	50.000
CEE	B.s	+	+	-*	-	-	-	-	-	-	-
EAA	B.s	+	+	+	+	+	-*	-	-	-	-
NBE	B.s	+	+	+	+	+	+	-*	-	-	-
AQE	B.s	+	+	+	+	+	+	-*	-	-	-
CEE	E.c	+	+	+	+	-*	-	-	-	-	-
EAA	E.c	+	+	+	+	+	-*	-	-	-	-

NBE	E.c	+	+	+	+	+	+	+	+	+	-*	-	-
CEE	P.e	+	+	+	+	+	+	-*	-	-	-	-	-
EAA	P.e	+	+	+	+	+	+	+	+	+	-*	-	-
NBE	P.e	+	+	+	+	+	+	+	+	+	-*	-	-
CEE	S.a	+	+	+	+	+	+	+	+	+	-*	-	-
EAA	S.a	+	+	+	+	+	+	+	+	+	-*	-	-
NBE	S.a	+	+	+	+	+	+	+	+	+	-*	-	-
CEE	S.t	+	+	-*	-	-	-	-	-	-	-	-	-
EAA	S.t	+	+	-*	-	-	-	-	-	-	-	-	-
NBE	S.t	+	+	+	+	+	+	-*	-	-	-	-	-
AQE	S.t	+	+	+	+	+	-*	-	-	-	-	-	-

Key: B.s = *Bacillus subtilis*, E.c = *Escherichia coli*, P.e = *Pseudomonas euroginosa*, S.a = *Staphylococcus aureus*, S.t = *Salmonella typhi*, C.a = *Candida albicans*, A.f = *Aspergillus flavus*, A.n = *Aspergillus niger*, CEE = crude ethanolic extract, EAA = ethyl acetate portion, NBE = n-butanol portion, AQE = residual aqueous portion, + = Turbidity observed, - = No turbidity observed, -* = MIC value.

Table 4: Minimum bactericidal concentrations (MBC) of the extracts *Chrozophora senegalensis*

Extracts	Organisms	Concentration (mg/ml)									
		0.098	0.195	0.390	0.780	1.563	3.125	6.250	12.500	25.000	50.000
CEE	B.s	+	+	+	φ	-	-	-	-	-	-
EAA	B.s	+	+	+	+	+	+	+	φ	-	-
NBE	B.s	+	+	+	+	+	+	+	φ	-	-
AQE	B.s	+	+	+	+	+	+	+	φ	-	-
CEE	E.c	+	+	+	+	+	+	+	+	φ	-
EAA	E.c	+	+	+	+	+	φ	-	-	-	-
NBE	E.c	+	+	+	+	+	+	+	φ	-	-
CEE	P.e	+	+	+	+	+	φ	-	-	-	-
EAA	P.e	+	+	+	+	+	+	+	φ	-	-
NBE	P.e	+	+	+	+	+	+	φ	-	-	-
CEE	S.a	+	+	+	+	+	+	+	+	+	φ
EAA	S.a	+	+	+	+	+	+	+	-*	-	-
NBE	S.a	+	+	+	+	+	+	+	-*	-	-
CEE	S.t	-	-	-	-	-	-	-	-	-	-
EAA	S.t	-	-	-	-	-	-	-	-	-	-
NBE	S.t	+	+	+	+	+	+	+	φ	-	-
AQE	S.t	+	+	+	+	φ	-	-	-	-	-

Key: B.s – *Bacillus subtilis*, E.c - *Escherichia coli*, P.e - *Pseudomonas euroginosa*, S.a - *Staphylococcus aureus*, S.t - *Salmonella typhi*, CEE - crude ethanolic extract, EAA - ethyl acetate portion, NBE - n-butanol portion, AQE - residual aqueous portion, + = growth, - = no growth, φ = MBC value.

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References

1. Ahmad, I.; Mehmood, Z. and Mohammed, F. (1998). Screening of Some India Medicinal Plants for their Antimicrobial Properties. *J Ethnopharmacol* **62**:182-193.
2. Bellomaria, B. and Kacou, P. (1995). Plants et Medicine Population d'Agboville (Cote D'Ivoire) *Fitoterapia* **LXVI**: 117 –141.
3. Bose, T.K. and Choudhary, K. (1991). *Tropical Garden Plants in Colour*. Horticulture and Allied Publishers. 27/3 Chakraberia lane, Calcutta, 700 020, India. pp. 1-10
4. Chung, K.T.; Thomson, W.R. and Wu-Yan, C.D. (1990). Growth Inhibition of Selected Food-Borne Bacteria by Plant Extracts. *J. Appl. Microbiol.* **71**: 398-401.
5. Clark, A.M. (1996). Natural Products as Resource for New Drugs. *Pharm. Res.* **13**:1133-1141.
6. Cordell, G.A. (2000). Biodiversity and Drug discovery – a Symbiotic Relationship. *Phytochem.* **55**: 463-480.
7. Dalziel, J.M. (1955). *The Useful plants of West Tropical Africa*. Crown Agents for Oversea Government and Administration 4, Millbank, London, S.W. 1. pp. 271-273.
8. Etkin, N.L. (1997). Antimalarial Plants used by Hausa in Northern Nigeria. *Tropical Doctor.* **27** 1: 12 – 16.
9. Harborne, J.B. (1993). *Phytochemistry*. Academic Press, London. pp. 89-131.
10. Hassan, M.M.; Oyewale, A.O.; Amupitan, J. O.; Abdullahi, M.S. and Okonkwo, E.M. (2004). Preliminary Phytochemical and antibacterial investigation of crude extracts of the root bark of *Detarium microcarpum*. *J. Chem. Soc. Nigeria* **29** 1: 26-29.
11. Lewis, W.H. and Elvin-Lewis, M.P.F. (1997). *Medical Botany-plants affecting Man's health*. John Wiley, New York, USA. pp. 36 – 55.
12. Odama, I.E.; Shok, M. and Olurinola, P.F. (1986). *Ceiba Petandra* (Silk Cotton Tree). Microbiological Evaluation of its Use in Traditional Wound Treatment. In: *The State of Medicinal Plant Research In Nigeria* (Sofowora, A. Ed.) Ile-Ife, Univ. Press. pp. 253-259.
13. Rahman, A-U. and Choudhary, M. I. (1999). Recent Studies on Bioactive Natural Products. *Pure App. Chem.* **71**(6) pp. 1079-1081.
14. Sidney, M.F.; William, J.M. and Elvyn, G.S. (1978). *Bailey and Scott's Diagnostic Microbiology*. C.V. Mosby: St. Louis. pp. 385-403.
15. Sofowora, A. (1993). Screening plants for Bioactive Agents. In: *Medicinal plants and Traditional medicine in Africa*. 2nd ed. Spectrum Books Ltd, Sunshine House, Ibadan. pp. 134-156.
16. Srinivasan, D.; Nathan, S.; Suresh, T. and Perumalsamy, P.Z. (2001). Antimicrobial Activity of Certain India Medicinal Plant used in Folkloric Medicine. *J. Ethnopharmacol.* **74**: 217-220.
17. Tignokpa, M.; Laurens, A.; Mboup, S. and Sylla, O. (1986). Popular Plants of Dakar Markets. *Int. J. Crude Drug Res.* **24** 2: 75-80.
18. Trease G.E. and Evans, W.C. (1978). *Pharmacognosy*. Baillier Tindal and Macmillan Publishers, London. pp. 256.
19. Trease, G. E. and Evans, W. C. (2002). *Pharmacognosy*. 15th Ed. Saunders Publishers, London. pp. 42 – 44, 221 – 229, 246 – 249, 304 – 306, 331 – 332, 391 – 393.
20. Vollekova, A.; Kost'álovà, D. and Sochorová, R. (2001). Isoquinoline Alkaloids from *Mahonia aquifolium* Stem bark is Active against *Malassezia spp.* *Folia Microbiol.* **46**:107-111.
21. Zwadyk, P. (1972). *Enteriobacteriaceae* in *Zinsser Microbiology*. 20th ed. George Thiene Verlag, Stuttgart. pp. 20 – 32.